

6c3e227f-03a1-4252-a1c1-fe1e8f49cf5f_51191_-_agung_putra...

 Unika Soegijapranata1

Document Details

Submission ID

trn:oid:::28973:111553042

19 Pages

Submission Date

Sep 10, 2025, 5:27 AM GMT+7

9,129 Words

Download Date

Sep 10, 2025, 5:41 AM GMT+7

50,208 Characters

File Name

6c3e227f-03a1-4252-a1c1-fe1e8f49cf5f_51191_-_agung_putra_v3.pdf

File Size

1.8 MB

10% Overall Similarity

The combined total of all matches, including overlapping sources, for each database.

Filtered from the Report

- Submitted works
- Internet sources
- Crossref database

Match Groups

-  **63** Not Cited or Quoted 9%
Matches with neither in-text citation nor quotation marks
-  **0** Missing Quotations 0%
Matches that are still very similar to source material
-  **4** Missing Citation 1%
Matches that have quotation marks, but no in-text citation
-  **0** Cited and Quoted 0%
Matches with in-text citation present, but no quotation marks

Top Sources

- 0%  Internet sources
- 10%  Publications
- 0%  Submitted works (Student Papers)

Integrity Flags

2 Integrity Flags for Review

-  **Replaced Characters**
95 suspect characters on 19 pages
Letters are swapped with similar characters from another alphabet.
-  **Hidden Text**
24 suspect characters on 2 pages
Text is altered to blend into the white background of the document.

Our system's algorithms look deeply at a document for any inconsistencies that would set it apart from a normal submission. If we notice something strange, we flag it for you to review.

A Flag is not necessarily an indicator of a problem. However, we'd recommend you focus your attention there for further review.

Match Groups

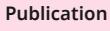
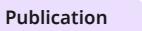
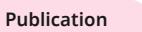
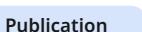
-  63 Not Cited or Quoted 9%
Matches with neither in-text citation nor quotation marks
-  0 Missing Quotations 0%
Matches that are still very similar to source material
-  4 Missing Citation 1%
Matches that have quotation marks, but no in-text citation
-  0 Cited and Quoted 0%
Matches with in-text citation present, but no quotation marks

Top Sources

- 0%  Internet sources
- 10%  Publications
- 0%  Submitted works (Student Papers)

Top Sources

The sources with the highest number of matches within the submission. Overlapping sources will not be displayed.

-   Miguel Ossandon, Ben Prickril, Avraham Rasooly. "Cancer Detection and Diagnosi... 2%
-   Widyaningsih, Wita. "Pengaruh Pemberian Sekretom sel Punca Mesenkimal Hipo... 1%
-   Metalia, Metalia. "Pengaruh gel Topikal Secretome Hypoxia Mesenchymal Stem ... <1%
-   Mayasari, Yunita Ika. "Pengaruh Secretom Hipoksia Mesenchymal Stem Cells Ter... <1%
-   Khawaja Husnain Haider. "Handbook of Regenerative Medicine - Stem Cell-Based ... <1%
-   Shiyi Zhang, Lin Jin, Xinyi Li, Jianhui Zhang, Ning Wang, Jianxiong Chen, Lingheng ... <1%
-   Octavianti, Yessi. "Pengaruh Pemberian Secretome Hypoxia Mesenchymal Stem C... <1%
-   Hagman, Karl. "SARS-CoV-2 Viremia in Patients Hospitalised for COVID-19", Karoli... <1%
-   Caramelo, Inês Isabel Nunes. "New Approaches on Hypoxic-Ischemic Encephalop... <1%
-   Kerry Layne. "100 Cases in Acute Medicine", CRC Press, 2019 <1%

11 Publication

Muthmainah, Hanik Badriyah Hidayati, Budi Yanti. "Improving Health for Better F... <1%

12 Publication

Tabish Qidwai. "Interaction of Coronavirus Disease 2019 with other Infectious an... <1%

13 Publication

Goodfellow, Samuel Melvin. "Investigation of Orthohantavirus Genetics in Roden... <1%

14 Publication

Victor R. Preedy. "Plant and Animal Proteins in Health and Disease Prevention", C... <1%

15 Publication

Xin Ma, Meng Li, Yuanyuan Zhang, Tingting Xu, Xinchen Zhou, Mengqi Qian, Zhire... <1%

16 Publication

Pachow, Collin. "Investigating the Roles of Microglia in Viral CNS Disease.", Unive... <1%

17 Publication

Deevia Bhana, Morten Skovdal, Kaymarlin Govender. "Young Masculinities and Se... <1%

18 Publication

Santos Bravo, Marta. "Antiviral Resistance and Persistent Replication of Cytomeg... <1%

19 Publication

Hamar, Ágoston László. "A COVID-19 Betegség Patomechanizmusának és Lefolyá... <1%

20 Publication

Lusiana, Dewi Masythah Darlan, Setyo Trisnasi, Agung Putra, Nur Dina Amalina, S... <1%

21 Publication

RC Sobti. "Biotechnological Innovation and Sustainable Developmental Goals - Fo... <1%

22 Publication

Yutong Zhao. "Lung Biology and Pathophysiology", CRC Press, 2024 <1%

23 Publication

Chen, Rui. "Investigating Cellular Dysfunction in Injury-Induced Neurological and... <1%

24 Publication

Ferreira, Victor Leonel Vieira. "SARS-CoV-2 Detection Methods: Where are we Now... <1%

25

Publication

Suvash C. Saha, Akshoy Ranjan Paul. "Biological Flow Modelling", CRC Press, 2025 <1%

26

Publication

Tjun Tang, Elizabeth O'Riordan, Stewart Walsh. "Cracking the Intercollegiate Gen... <1%

27

Publication

Genter, Franziska. "Investigating Groundwater Self-Supply as a Safely Managed ... <1%

28

Publication

Sebastian L. Johnston, Paul O'Byrne. "Exacerbations of Asthma", CRC Press, 2019 <1%

29

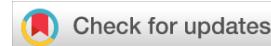
Publication

Haque, Nazmul. "A Novel Process Using Human Sera for the Production of Secret... <1%

30

Publication

Rehman, Tanzeel Ur. "An In-Depth Study of DOPE:DOPC Liposomes to Maximize T... <1%



CLINICAL PRACTICE ARTICLE

4 **REVISED Case series of the first three severe COVID-19 patients****treated with the secretome of hypoxia-mesenchymal stem cells in Indonesia**

[version 3; peer review: 2 approved]

Agung Putra ¹⁻³, Agus Widyatmoko⁴, Sugeng Ibrahim^{1,5}, Fajar Amansyah⁶, Farid Amansyah⁶, Mukti Arja Berlian⁷, Retnaningsih Retnaningsih ⁸, Zenitalia Pasongka⁹, Flora Eka Sari¹⁰, Basuki Rachmad¹¹

¹Stem Cell and Cancer Research (SCCR), Faculty of Medicine, Universitas Islam Sultan Agung (Unissula), Semarang, Central Java, Indonesia

²Department of Postgraduate Biomedical Science, Faculty of Medicine, Universitas Islam Sultan Agung (Unissula), Semarang, Central Java, Indonesia

³Department of Pathological Anatomy, Faculty of Medicine, Universitas Islam Sultan Agung (Unissula), Semarang, Central Java, Indonesia

⁴Department of Internal Medicine, Faculty of Medicine, Universitas Muhammadiyah Yogyakarta, Yogyakarta, Yogyakarta, Indonesia

⁵Department of Molecular Biology, Faculty of Medicine, Universitas Katholik (Unika) Soegijapranata, Semarang, Central Java, Indonesia

⁶Department of Internal Medicine, Bhayangkara Hospital, Makassar, South Sulawesi, Indonesia

⁷Department of Internal Medicine, Dr. Esnawan Antariksa Air Force Hospital, Jakarta, Jakarta, Indonesia

⁸Department of Neurology and Intensive Care Unit, Kariadi Hospital, Universitas Diponegoro, Semarang, Central Java, Indonesia

⁹Faculty of Medicine, Universitas Udayana, Denpasar, Bali, Indonesia

¹⁰Department of Pulmonary Medicine, Dr. Esnawan Antariksa Air Force Hospital, Jakarta, Jakarta, Indonesia

¹¹Department of Intensive Care Unit, Gatot Soebroto Army Hospital, Jakarta, Jakarta, Indonesia

V3 First published: 22 Mar 2021, 10:228

<https://doi.org/10.12688/f1000research.51191.1>

Second version: 22 Jul 2021, 10:228

<https://doi.org/10.12688/f1000research.51191.2>

Latest published: 22 Oct 2021, 10:228

<https://doi.org/10.12688/f1000research.51191.3>

Abstract

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is responsible for the outbreak of coronavirus disease 2019 (COVID-19), which has been rapidly spreading. Several guideline therapies have been proposed as a possible treatment for SARS-CoV-2, however, these therapies are not sufficient to treat a severe condition of SARS-CoV-2 infection characterised by the increase of D-dimer and C-reactive protein (CRP) levels, and patchy ground-glass opacities (GGOs). Secretome-mesenchymal stem cells (S-MSCs) produced by MSCs under hypoxia could excessively release several anti-inflammatory cytokines and growth factors to control the COVID-19 cytokine storm and accelerate lung injury improvement. This is the first study investigating the clinical outcomes of three severe COVID-19 patients admitted to the intensive care unit of three different

Open Peer Review**Approval Status**

1

2

version 3

(revision)
22 Oct 2021

[view](#)**version 2**

(revision)
22 Jul 2021

[view](#)**version 1**

22 Mar 2021

[view](#)[view](#)

1. Rajesh Ramasamy , Universiti Putra

hospitals in Indonesia treated with S-MSCs. The decrease of D-dimer and CRP level was reported for all patients treated with S-MSCs. This was in line with improvement of pulmonary radiology, blood gas level, and hematologic assessment. In conclusion, these cases suggest that S-MSCs could effectively control D-dimer, CRP level and GGOs of severe COVID-19 patients associated with recovered pulmonary function.

Keywords

COVID-19, secretome, mesenchymal stem cells

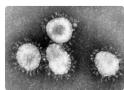
Malaysia, Selangor, Malaysia

2. Guido Moll , Berlin Institute of Health, Charité - University Medicine Berlin, Berlin, Germany

Any reports and responses or comments on the article can be found at the end of the article.



This article is included in the **Emerging Diseases and Outbreaks** gateway.



This article is included in the **Coronavirus (COVID-19)** collection.

Corresponding author: Agung Putra (dr.agungptr@gmail.com)

Author roles: Putra A: Conceptualization, Validation, Writing – Review & Editing; Widyatmoko A: Conceptualization, Data Curation, Investigation; Ibrahim S: Formal Analysis, Resources; Amansyah F: Methodology, Resources, Visualization; Amansyah F: Conceptualization, Data Curation, Resources; Berlian MA: Conceptualization, Resources, Supervision; Retnaningsih R: Formal Analysis, Visualization; Pasongka Z: Investigation, Project Administration; Sari FE: Writing – Original Draft Preparation; Rachmad B: Resources, Writing – Original Draft Preparation

Competing interests: No competing interests were disclosed.

Grant information: This study was funded by the Provincial Government of Central Java, Indonesia.

The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Copyright: © 2021 Putra A *et al.* This is an open access article distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

How to cite this article: Putra A, Widyatmoko A, Ibrahim S *et al.* **Case series of the first three severe COVID-19 patients treated with the secretome of hypoxia-mesenchymal stem cells in Indonesia** [version 3; peer review: 2 approved] F1000Research 2021, 10:228 <https://doi.org/10.12688/f1000research.51191.3>

First published: 22 Mar 2021, 10:228 <https://doi.org/10.12688/f1000research.51191.1>

REVISED Amendments from Version 2

The new version of our article provides 2 additions regarding the clarification of chondrogenic differentiation assessment and the update of isotype staining of MSCs marker analysis using flow cytometry.

Any further responses from the reviewers can be found at the end of the article

Introduction

Since December 2019, severe acute respiratory syndrome corona virus 2 (SARS-CoV-2), responsible for the outbreak of coronavirus disease 2019 (COVID-19), has been rapidly spreading worldwide¹. The number of infected persons has exceeded 87 million with 2 million deaths globally². Several guideline therapies such as remdesivir and convalescent plasma have been proposed as possible treatment for SARS-CoV-2, however these treatments remain controversial. Moreover, these therapies were not effective to treat severe infection of SARS-CoV-2 due to these treatments potentially inducing the robust cytokine storm^{3,4}. A previous study demonstrated that there is a correlation between disease severity and the release of proinflammatory cytokines, such as tumor necrosis factor- α (TNF- α), IL-6, IL-1B, IL-4, IFN- γ , IFN- γ -induced protein 10 (IP10), monocyte chemoattractant protein-1 (MCP-1), macrophage inflammatory protein-1a (MIP-1a), and granulocyte-colony stimulating factor (G-CSF)⁵. This finding was confirmed by the high plasma cytokines found in the most severe COVID-19 patients associated with extensive lung damage^{6,7}. Therefore, finding an effective therapeutic option to hamper the devastating cytokine storm of COVID-19 and regenerate the damaged lung is crucial. Previous studies recently reported several benefits of mesenchymal stem cells (MSCs) under hypoxia condition to inhibit robust proinflammatory cytokines and repair extensive tissue damage by releasing several anti-inflammatory cytokines and growth factors⁸.

The use of hypoxia-MSCs (H-MSCs) could become an alternative solution to treat the severe cytokine storm of COVID-19. A previous study reported that hypoxia precondition treated on MSCs (H-MSCs) could enhance their survival to reach the damaged area⁹. However, blood clots appearing during

the severe phase of COVID-19 could block the H-MSCs trajectory into the damaged area¹⁰. Other studies reported that H-MSCs could enhance the release of their active soluble molecules, known as secretome-MSCs (S-MSCs) such as IL-10, TGF beta, VEGF and PDGF, which are useful in hampering inflammation and improving tissue healing¹¹. Therefore, isolating and concentrating the exact active soluble molecule of S-MSCs is a possible strategy to control the cytokine storm of COVID-19, and, in addition, to accelerate the damaged lung improvement.

In a recent study, we successfully isolate S-MSCs from their culture medium using tangential flow filtration (TFF) strategy with several molecular weight cut-off category¹². In this Clinical Practice article, we report on three severe COVID-19 patients with several comorbidities who were treated with S-MSCs in three different hospitals in Indonesia. This is the first report to describe the complete monitoring of these three patients.

Ethical considerations

Ethical clearance for the use of S-MSCs in COVID-19 cases and the protocol for administration was obtained from the Health Research Ethics Committee of Bethesda Hospital, Yogyakarta, Indonesia (approval number, No.91/KEPK-RSB/VI/20). Written informed consent for treatment with S-MSCs was obtained from each patient prior to treatment. All patients were treated with standard treatments for severe condition of COVID-19 infection, in addition to novel S-MSCs therapy regarding. Each patient was treated with three, four and six doses of 1 mL S-MSCs every 12 h (with molecular weight cut-off combination of 10–50 kDa 50%, 50–100 kDa 25%, and 100–300 kDa 25%) via deltoid intramuscular injection, respectively. A different S-MSCs dose was utilized in the three patients due to the preliminary nature of this treatment.

Case reports**Case no. 1**

A 54-year-old Indonesian male with severe hypertension was diagnosed with COVID-19 on August 28, 2020 and admitted to Dr. Esnawan Antariksa Air Force Hospital, Jakarta, Indonesia intensive care unit (ICU) with cough and dyspnea (Table 1) and was treated with standard treatment (Table 2). The examination showed a temperature of 36°C, a 102/min pulse, a respiratory rate of 32/min, and a blood pressure

Table 1. Clinical characteristics of COVID-19 patients receiving S-MSCs.

Patient no.	Sex	Age	Clinical classification	Days of S-MSCs therapy from symptom onset	Principal symptoms	Comorbidity
1	M	54	Severe	11	Cough, dyspnea	Severe hypertension
2	M	53	Severe	5	Cough, dyspnea, chest pain and fatigue	Type-2 diabetes mellitus
3	M	72	Severe	8	Abdominal pain, diarrhea, anosmia, cough and sore	Mild hypertension, liver failure, long-term sequelae of stroke, thalassemia minor

of 200/100. Blood gas analysis showed decreased oxyhemoglobin saturation (SO_2 , 80.6%; normal reference: 95–100%), CO_2 partial pressure (PCO_2 , 22.9 mmHg; normal reference: 38–42 mmHg), oxygen partial pressure (PO_2 , 37.6 mmHg; normal reference: 70–99 mmHg) and HCO_3^- (18.9 mmol/L; normal reference: 22–29 mmol/L). Laboratory studies showed increased white blood cells (WBC) count ($17.2 \times 10^9/\text{L}$; normal reference: $4–10 \times 10^9/\text{L}$), monocyte count (9%; normal reference: 2–8%) and decreased lymphocyte count (15%; normal reference: 20–40%). On August 29, D-dimer was increased (1540 ng/mL; normal reference: 0–231 ng/mL) in line with the elevation of C-reactive protein (CRP, 61.7 mg/dL; normal reference: 0–8.1 mg/L) (Table 3). Chest X-ray showed bronchopneumonia with bilateral ground glass opacities (GGOs) and cardiomegaly condition (Figure 1).

The patient was treated with 1 mL S-MSCs three times every 12 h via deltoid intramuscular on August 30 and 31.

On September 4, SO_2 was increased (99.6%) with increased PCO_2 (36.2 mmHg), PO_2 (198.7 mmHg), and HCO_3^- (24.7 mmol/L). Laboratory studies showed normal WBC count ($7.4 \times 10^9/\text{L}$), monocyte count (5%) and lymphocyte count (20%). D-dimer and CRP level were decreased (1297 ng/mL and 2.33 mg/dL, respectively) (Table 3). The chest X-ray still showed a bronchopneumonia with cardiomegaly condition (Figure 1). On September 10, the patient was reported negative from

COVID-19 infection. The chest X-ray showed improvement with no both bronchopneumonia and cardiomegaly observed. D-dimer was decreased (384 ng/mL), and CRP was normal (0.31 mg/dL). On 20 September, the patient has no cough and dyspnea. The examination showed that SO_2 was 98.7%. He was discharged from ICU and mobilized into rehabilitation room for standard recovery of physical activity.

Case no. 2

A 53-year-old Indonesian male with type 2 diabetes mellitus was diagnosed with COVID-19 on November 16, 2020. He was admitted to Gatot Soebroto Army Hospital, Jakarta, Indonesia on November 17 with cough, dyspnea, chest pain and fatigue (Table 1). On November 19, he was admitted to the ICU due to worsened dyspnea and treated with standard treatment (Table 2). Chest X-ray revealed cardiomegaly with lung edema, bilateral GGOs, aorta elongation and aorta atherosclerosis (Figure 2). Blood gas analysis showed PCO_2 (29.9 mmHg), PO_2 (177.1 mmHg) and HCO_3^- (20.8 mmol/L), however SO_2 was still normal (98%). On November 21, the SO_2 was 90.6% (abnormal) with reduced PO_2 (57.5 mmHg). Laboratory studies showed normal WBC count ($6.02 \times 10^9/\text{L}$) with increased neutrophil count (81%, normal reference: 50–70%), monocyte count (9%) and decreased lymphocyte count (10%). D-dimer was abnormal (880 ng/mL). Blood chemistries revealed elevations in CRP (160 mg/L) and fasting plasma glucose (FPG, 398 mg/dL; normal reference: 70–140 mg/dL) (Table 3).

Table 2. Standard treatment received by the three patients receiving S-MSCs.

Patient no.	Drugs Administrated			Oxygen support	
	Antiviral treatment	Antibiotic or antifungal treatment	Corticosteroids treatment	Before S-MSCs Therapy	After S-MSCs therapy
1	Remdesivir 100 mg, q24h i.v. in 5 days	Azithromycin po. and Levofloxacin i.v.	Dexamethasone i.v.	High-flow nasal oxygen	nasal cannula
2	Oseltamivir 75 mg, q12h po. in 5 days	Azithromycin po. Levofloxacin i.v.	None	High-flow nasal oxygen	nasal cannula
3	Favipiravir 600mg q12h po. In 5 days	Azithromycin po., Tazobactam Sodium and Levofloxacin i.v.	Dexamethasone i.v.	High-flow nasal oxygen	nasal cannula

po., per os; i.v., i.v. injection; q12h, every 12 h; q24h, every 24 h.

Table 3. Comparison of laboratory parameters before and after S-MSCs treatment.

Clinical Factors	Before S-MSCs treatment			After S-MSCs treatment		
	Patient 1	Patient 2	Patient 3	Patient 1	Patient 2	Patient 3
CRP (mg/L, normal reference: 0–8.1 mg/L)	61.7	160	118	0.37	5.12	8.5
D-dimer (ng/mL, normal reference: 0–231 ng/mL)	1540	880	235	384	660	87
Lymphocyte (%), normal reference: 20–40%)	15	10	11.7	20	27	25.8
SO_2 (%), normal reference: 95–100%)	80.6	90.6	85	99.6	95.9	98

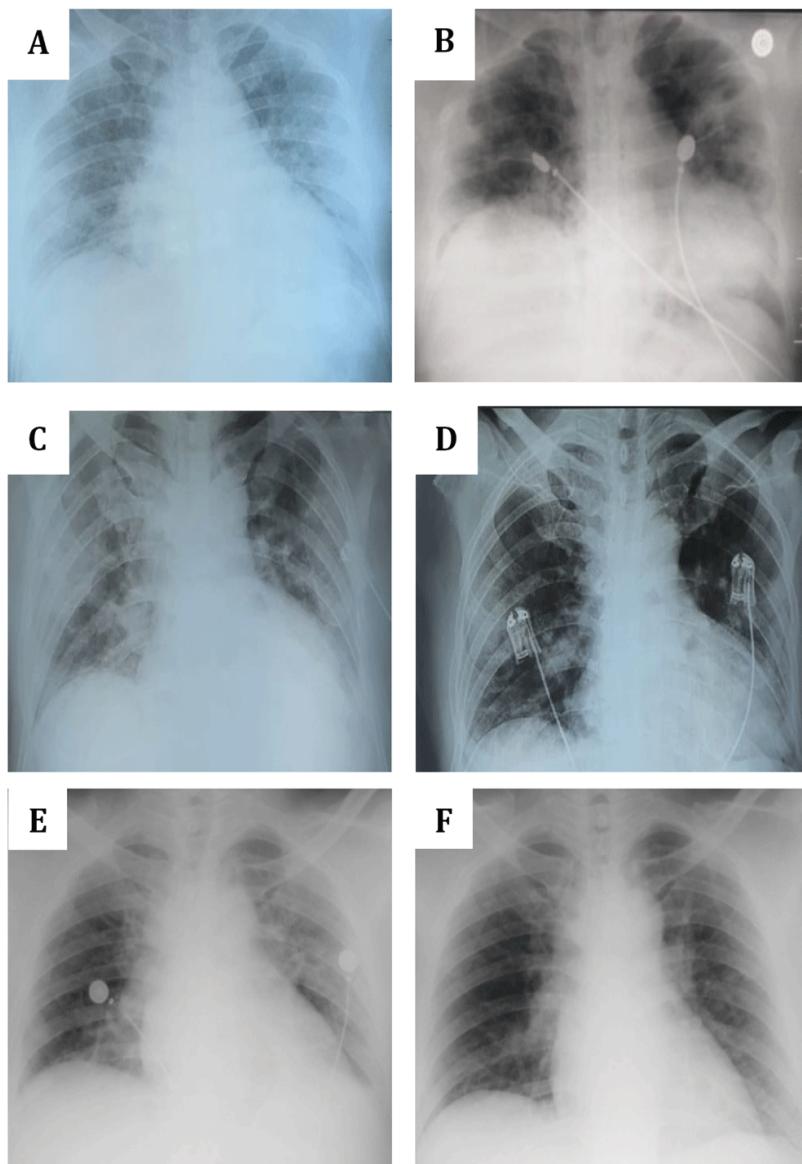


Figure 1. Chest X-ray of the three patients. (A and B) Patient 1. (A) August 29 (a day post onset of illness (dpoi)), showing bronchopneumonia with bilateral ground glass opacities (GGOs) and cardiomegaly; (B) September 10 (11 dpoi), showing the absorption of bilateral GGOs with no bronchopneumonia and cardiomegaly. (C and D) Patient 2. (C) November 19 (3 dpoi), showing cardiomegaly with lung edema, bilateral GGOs aorta elongation and aorta atherosclerosis; (D) November 22 (6 dpoi), showing improvement with minimum infiltrate on pulmonalis dextra and sinistra. (E and F) Patient 3. (C) December 23 (7 dpoi), showing worsened bilateral GGOs with cardiomegaly and aortic atherosclerosis; (F) December 28 (12 dpoi), showing decreased bilateral GGOs, cardiomegaly and aortic elongation.

Due to the patient's worsening condition, he was injected with 1 mL S-MSCs four times every 12 h via deltoid intra-muscular injection on November 21 and 22. On the same day, 6 hours after injection, SO_2 increased (98.7%) with increased PO_2 (138.5 mmHg), pH (7.509) and HCO_3^- (29.7 mmol/L). On November 22, chest X-ray showed improvement with minimum infiltrate on pulmonalis dextra and sinistra (Figure 1). On November 23, oxygen saturation was normal (95.9%), with normal PCO_2 (36.1 mmHg) and PO_2 (71.5 mmHg). Laboratory studies showed increased neutrophil count (85%)

with normal monocyte count (6%) and decreased lymphocyte count (9%). D-dimer was decreased (660 ng/mL). Blood chemistries revealed decreased fasting plasma glucose (277 mg/dL) and CRP (5.12 mg/dL (Table 3). On December 5, the patient has no cough, dyspnea, chest pain and fatigue. Examination showed that SO_2 was 99.2%. He was discharged from ICU and mobilized into rehabilitation room for physical activity recovery. On December 28, laboratory studies showed normal neutrophil (57%) and lymphocyte count (27%) with increased monocyte count (11%). Chest X-ray showed

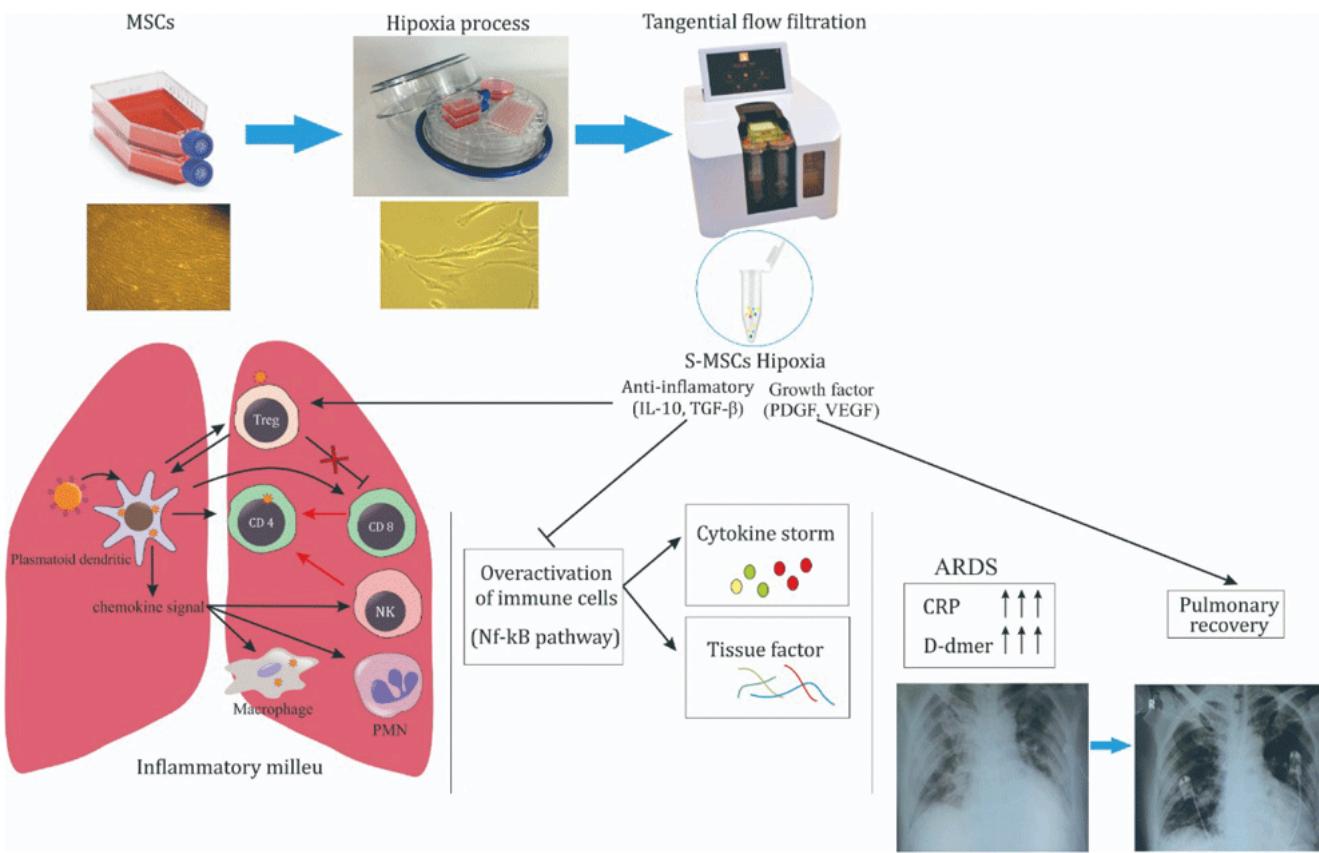


Figure 2. Schematic of cytokine storm on severe COVID-19 treated by S-MSCs. MSCs were incubated in 5% O₂ hypoxia condition and S-MSCs was isolated from the culture medium using TFF technique¹². S-MSCs contain soluble molecules, including IL-10, TGF-β, PDGF and VEGF. IL-10 and TGF-β inhibit NF-κB pathways activation of overactivated immune cells leading to cytokine storm inhibition, characterized by the decreased level of CRP and D-dimer. Under controlled inflammatory milieu, VEGF and PDGF promote the improvement of bilateral GGOs marked by pulmonary recovery.

normal cardiac physiology and no infiltrate or nodule in both pulmonalis.

Case no. 3

A 72-year-old Indonesian male with mild hypertension, liver failure, long-term sequelae of stroke and thalassemia minor was diagnosed with COVID-19 on December 16, 2020 and admitted to Bhayangkara Hospital, Makassar, Indonesia with abdominal pain, diarrhea, anosmia, cough and sore throat in the last three days (Table 1). The examination showed a temperature of 37.3°C, pulse of 82/min, respiratory rate of 24/min, blood pressure of 140/90 mmHg, and SO₂ of 97%. Laboratory studies showed a decreased WBC count (3.1×10^9 /L), with normal neutrophil count (53.4%), lymphocyte count (26.8%) and increased monocyte count (18.8%). Chest X-ray showed bilateral GGOs. On December 23, the patient's dyspnea worsened and he was admitted to the ICU and treated with standard treatment (Table 2). Blood gas analysis revealed that oxygen saturation was decreased (85%) with decreased PCO₂ (30 mmHg), PO₂ (49 mmHg) and HCO₃⁻ (19 mmol/L). Laboratory tests showed an increased neutrophil count (72.9%), monocyte count (14.9%) and decreased lymphocyte count (11.7%).

D-dimer was increased (235 ng/mL) and blood chemistries revealed elevations in CRP (118 mg/L (Table 3)). Chest X-ray showed worsened bilateral GGOs with cardiomegaly and aortic atherosclerosis (Figure 1).

On December 24–26, the patient was treated with 1 mL S-MSCs six times every 12 h via deltoid intramuscular injection in addition to other standard treatment (Table 2). The day after the first S-MSC injection, oxygen saturation increased (98%). The examination showed a temperature of 37°C, pulse of 90/min, respiratory rate of 28/min and blood pressure of 120/80 mmHg. On December 28, examination showed a temperature of 37°C, pulse of 80/min, respiratory rate of 24/min and blood pressure of 120/80 mmHg. Oxygen saturation was normal (98%). Laboratory studies showed normal neutrophil count (78.3%) monocyte count (7.2%) and lymphocyte count (25.8%). D-dimer was decreased (86.9 ng/mL). Blood chemistries revealed decreased CRP (8.5 mg/dL (Table 2)). Chest X-ray showed decreased bilateral GGOs, cardiomegaly and aortic elongation (Figure 1). On December 30, the patient has no abdominal pain, diarrhea, anosmia, cough and sore throat. The examination showed that SO₂ was 99%. He was

discharged from ICU and mobilized into rehabilitation room for standard physical activity recovery. On January 6, the patient was negative for COVID-19.

Discussion

Severe pneumonia COVID-19 is characterized by rapid viral infection, excessive inflammatory cell infiltration and robust cytokine storm associated with an increase of D-dimer and CRP levels, resulting in acute respiratory distress syndrome (ARDS)¹³. A previous study reported that the cytokine storm of COVID-19 was associated with an increase of several proinflammatory cytokines, including TNF- α , IL-1, IL-6, IL-17A and granulocyte macrophage colony-stimulating factor (GM-CSF) in the plasma. In line with this phenomenon, lymphocyte count in severe COVID-19 patients' peripheral blood was decreased^{14,15}. Another study also revealed that lymphocyte count in severe COVID-19 patients was reduced due to overactivated immune cells, known as macrophage activated syndrome (MAS). The MAS potentially promotes the excessive cytokines storm characterized by the increase of CRP and D-dimer level leading to bilateral GGOs¹⁶. S-MSCs contain several anti-inflammatory cytokines, including IL-10 and TGF- β , to control the overactivated immunity and hamper the storm's excessive cytokines. In addition, S-MSCs also have several growth factors, such as VEGF and PDGF, that could accelerate lung injury improvement in COVID-19 patient^{8,11}.

MSCs were isolated from umbilical cord and exhibited typical monolayers of spindle-shaped fibroblast-like cells with plastic adherent capability under standard culture condition. The cultured MSCs have a high level of surface antigens, including CD90, CD73 and CD105, and lacked the expression CD34, CD45, CD11b, CD19, and HLA-DR, represented as Lin. On the other hand, they also showed the capability to differentiate into osteocyte and adipocyte (see extended data, figure S1 for more details). To induce hypoxia condition, the MSCs were treated under 5% O₂ condition in a hypoxia incubation chamber for 24 h. After incubation, the culture medium of H-MSCs was collected and filtered using TFF strategy with molecular weight cut-off of 10–50 kDa, 50–100 kDa, and 100–300 kDa to produce the S-MSCs (Figure 2). This strategy was employed to obtain the desired cytokines and growth factors in that range, such as IL-10, TGF- β , VEGF and PDGF. In this study, we used S-MSCs with the combination of molecular weight cut-off of 10–50 kDa (50%), 50–100 kDa (25%), and 100–300 kDa (25%). The analysis of cytokines and growth factors level on this S-MSCs combination has been employed using flow cytometry CBA detection and showed a feasible result (see extended data, table S1 for more details).

This is the first report that suggests the feasibility of S-MSC therapy of three severe COVID-19 patients in Indonesia. All patients showed bilateral GGOs on chest X-ray before treatment. The hematologic findings showed elevated D-dimer and CRP level in addition to lymphocytopenia and decrease of SO₂ and PO₂. These patients received several doses of S-MSCs (1 mL/dose) / 12 h which results in favorable outcomes. This study showed that between three and six doses of S-MSCs were well tolerated by the patients. The clinical symptoms were

significantly improved with patchy GGO improvement, associated with the decrease of D-dimer and CRP level and increase of SO₂ and PO₂. From our observation, we would suggest that six doses of S-MSCs performed the most optimal treatment in the COVID-19 patients. These results suggest that the immune system's excessive inflammation and overactivation were alleviated by anti-inflammatory cytokines contained in S-MSCs, while the high level of growth factors in S-MSCs could also accelerate the improvement of GGOs. Based on our preliminary results, S-MSC therapy could be a promising and safe rescue option to treat severe COVID-19 patients.

The first key factor associated with effective S-MSC therapy is the controlled immune system overactivation resulting in the alleviated cytokines storm characterized by the decrease in CRP and D-dimer levels. In our observations, the level of CRP and D-dimer was decreased in all patients treated with S-MSCs. This data suggest that S-MSCs could effectively control the overactivated immune cells. A previous study reported that S-MSCs could control proinflammatory immune cells due to their anti-inflammatory cytokines, such as IL-10 and TGF- β ^{11,17}. IL-10 could hamper the inflammatory cells' activity by activating tyrosine kinase-2 and Janus tyrosine kinase 1 (JAK1) and inhibiting NF- κ B pathways leading to the decreased expression of proinflammatory cytokines such as TNF- α , IL-1, IL-6 and IL-17A^{9,18}. On the other hand, TGF- β could activate the regulatory subset of T lymphocyte (Treg) by initiating the FoxP3 expression, resulting in suppressing overactivated immune cells¹⁹ (Figure 2).

Another key factor associated with the efficacy of S-MSCs treatment is the acceleration of lung injury improvement characterized by the improvement pulmonary radiological results, decrease of CRP and D-dimer level resulting in the controlled inflammation leading to normal pulmonary function marked by normal SO₂. In our patients, decreased CRP and D-dimer was associated with the improvement of patchy GGOs and normal SO₂ in all patients treated with S-MSCs. These data suggest that along with the controlled immune overactivation, the growth factors contained in S-MSCs could rapidly improve lung regeneration post excessive cytokine storm exposure. A previous study revealed that S-MSCs could accelerate wound healing due to their growth factors, particularly VEGF and PDGF²⁰. VEGF and PDGF could accelerate the repair of leaky pulmonary blood vessels and accelerate lung injury improvement through the MEK and Akt pathway resulting in the acceleration of angiogenesis and reduced pulmonary infiltrate characterized by decreased bilateral GGOs^{21,22} (Figure 2).

In our patients, no severe adverse effects were observed. However, the limitation of our report is that the dynamic changes of cytokine and growth factors during treatment were not investigated. On the other side, we also did not perform the chondrogenic differentiation analysis of MSCs. Nevertheless, the preliminary result seems promising.

In conclusion, we report that S-MSC therapy shows a potential therapeutic effect and low risk in severe COVID-19

19 patient treatment. We observed that between three and six doses of S-MSCs, with weight cut-off combination of 10–50 kDa (50%), 50–100 kDa (25%) and 100–300 kDa (25%), rapidly controlled the excessive cytokine storm in our patients and improved lung injury. Six doses showed optimum outcome. The treatment time point and the clear clinical advantages of S-MSCs therapy need to be further investigated in randomized clinical studies.

Consent

Written informed consent for publication of this case report, along with any associated images, was obtained from all three patients.

Written informed consent was also obtained from the patients to undergo treatment using the novel treatment.

References

1. Munster VJ, Koopmans M, van Doremalen N, et al.: **A Novel Coronavirus Emerging in China - Key Questions for Impact Assessment.** *N Engl J Med.* 2020; **382**(8): 692-694.
[PubMed Abstract](#) | [Publisher Full Text](#)
2. Johns Hopkins University & Medicine: **Coronavirus Resource Center.** 2021; Accessed January 22,2021.
[Reference Source](#)
3. Dubert M, Visseaux B, Isernia V, et al.: **Case report study of the first five COVID-19 patients treated with remdesivir in France.** *Int J Infect Dis.* 2020; **98:** 290-293.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
4. Agarwal A, Mukherjee A, Kumar G, et al.: **Convalescent plasma in the management of moderate covid-19 in adults in India: open label phase II multicentre randomised controlled trial (PLACID Trial).** *BMJ.* 2020; **371:** m3939.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
5. Johnson BS, Laloraya M: **A cytokine super cyclone in COVID-19 patients with risk factors: the therapeutic potential of BCG immunization.** *Cytokine Growth Factor Rev.* 2020; **54:** 32-42.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
6. Huang C, Wang Y, Li X, et al.: **Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China.** *Lancet.* 2020; **395**(10223): 497-506.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
7. Yang X, Yu Y, Xu J, et al.: **Clinical course and outcomes of critically ill patients with SARS-CoV-2 pneumonia in Wuhan, China: a single-centered, retrospective, observational study.** *Lancet Respir Med.* 2020; **8**(5): 475-481.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
8. Eleuteri S, Fierabracci A: **Insights into the Secretome of Mesenchymal Stem Cells and Its Potential Applications.** *Int J Mol Sci.* 2019; **20**(18): 4597.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
9. Muhar AM, Putra A, Warli SM, et al.: **Hypoxia-Mesenchymal Stem Cells Inhibit Intra-Peritoneal Adhesions Formation by Upregulation of the IL-10 Expression.** *Open Access Maced J Med Sci.* 2019; **7**(23): 3937-3943.
[PubMed Abstract](#) | [Free Full Text](#)
10. Can A, Coskun H: **The rationale of using mesenchymal stem cells in patients with COVID-19-related acute respiratory distress syndrome: What to expect.** *Stem Cells Transl Med.* 2020; **9**(11): 1287-1302.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
11. Chen L, Xu Y, Zhao J, et al.: **Conditioned medium from hypoxic bone marrow-derived mesenchymal stem cells enhances wound healing in mice.** *PLoS One.* 2014; **9**(4): e96161.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
12. Harasztí RA, Miller R, Stoppato M, et al.: **Exosomes Produced from 3D Cultures of MSCs by Tangential Flow Filtration Show Higher Yield and Improved Activity.** *Mol Ther.* 2018; **26**(12): 2838-2847.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
13. Channappanavar R, Perlman S: **Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology.** *Semin Immunopathol.* 2017; **39**(5): 529-539.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
14. Chen N, Zhou M, Dong X, et al.: **Epidemiological and clinical characteristics of 99 cases of 2019 novel coronavirus pneumonia in Wuhan, China: a descriptive study.** *Lancet.* 2020; **395**(10223): 507-513.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
15. Sun HB, Zhang YM, Huang LG, et al.: **The changes of the peripheral CD4+ lymphocytes and inflammatory cytokines in Patients with COVID-19.** *PLoS One.* 2020; **15**(9): e0239532.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
16. Otsuka R, Seino KI: **Macrophage activation syndrome and COVID-19.** *Inflamm Regen.* 2020; **40:** 19.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
17. Putra A, Ridwan FB, Putridewi AI, et al.: **The Role of TNF-α induced MSCs on Suppressive Inflammation by Increasing TGF-β and IL-10.** *Open Access Maced J Med Sci.* 2018; **6**(10): 1779-1783.
[PubMed Abstract](#) | [Free Full Text](#)
18. Darlan DM, Munir D, Putra A, et al.: **MSCs-released TGFβ 1 generate CD4+ CD25+ Foxp3+ in T-reg cells of human SLE PBMC.** *J Formos Med Assoc.* 2021; **120**(1 Pt 3): 602-608.
[PubMed Abstract](#) | [Publisher Full Text](#)
19. Ahangar P, Mills SJ, Cowin AJ: **Mesenchymal Stem Cell Secretome as an Emerging Cell-Free Alternative for Improving Wound Repair.** *Int J Mol Sci.* 2020; **21**(19): 7038.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
20. Fournier NM, Lee B, Banasr M, et al.: **Vascular endothelial growth factor regulates adult hippocampal cell proliferation through MEK/ERK- and PI3K/Akt-dependent signaling.** *Neuropharmacology.* 2012; **63**(4): 642-52.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
21. Fang J, Huang X, Han X, et al.: **Endothelial progenitor cells promote viability and nerve regenerative ability of mesenchymal stem cells through PDGF-BB/PDGFR-β signaling.** *Aging (Albany NY).* 2020; **12**(1): 106-121.
[PubMed Abstract](#) | [Publisher Full Text](#) | [Free Full Text](#)
22. Dhingra S, Sharma AK, Arora RC, et al.: **IL-10 attenuates TNF-alpha-induced NF kappaB pathway activation and cardiomyocyte apoptosis.** *Cardiovasc Res.* 2009; **82**(1): 59-66.
[PubMed Abstract](#) | [Publisher Full Text](#)

Data availability

Extended data

Open Science Framework: Secretome H-MSCs for COVID-19 extended data. <https://doi.org/10.17605/OSF.IO/RJ8PC>

Data are available [under the terms of the Creative Commons Zero “No rights reserved” data waiver \(CC0 1.0 Public domain dedication\).](#)

Acknowledgements

We would like to thank Gatot Soebroto Army Hospital, Dr. Esnawan Antariksa Air Force Hospital Jakarta and Bhayangkara Hospital, Makassar, Indonesia for supporting the medical data collection. We would also thank to Stem Cell and Cancer Research (SCCR) laboratory for supporting this study.

Open Peer Review

Current Peer Review Status: ✓ ✓

Version 3

Reviewer Report 16 March 2022

<https://doi.org/10.5256/f1000research.78517.r97656>

1 © 2022 Ramasamy R. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

✓ **Rajesh Ramasamy** 

Department of Pathology, Universiti Putra Malaysia, Selangor, Malaysia

21 The authors have provided clarifications regarding the requested technical details. Although the reported clinical research is not a comprehensive clinical trial, the use of MSCs in 3 clinical cases brought insight to further exploring the mesenchymal stem cell therapy for severe lung-injured COVID-19 patients.

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Mesenchymal Stem Cell and Immunomodulation

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Version 2

Reviewer Report 10 September 2021

<https://doi.org/10.5256/f1000research.58709.r90165>

1 © 2021 Ramasamy R. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

? **Rajesh Ramasamy** 

¹ Department of Pathology, Universiti Putra Malaysia, Selangor, Malaysia

² Department of Pathology, Universiti Putra Malaysia, Selangor, Malaysia

The authors have submitted an acceptable characterisation of MSCs; however, the characterisation data that include immunophenotyping and mesodermal differentiation would be complete when:

1. chondrogenic differentiation data is included. A complete mesodermal differentiation based on acceptable standards comprises osteo, adipose and chondro differentiations;
2. it has provided isotype or unstained control for lineage negative staining. As a practice, the negative population is usually located within the first quarter or first half of the antibody axis. In the current supplementary data, the negative population has been depicted in the centre of the antibody axis.

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Mesenchymal Stem Cell and Immunomodulation

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 02 Oct 2021

Agung Putra

Dear Reviewer:

2 Thank you for reviewing our manuscripts, entitled "Case series of the first three severe COVID-19 patients treated with the secretome of hypoxia-mesenchymal stem cells in Indonesia". We appreciate the time and effort that reviewers dedicated to providing some feedback on our manuscript. We have incorporated most of the suggestions made by the reviewers.

Reviewer Comments:

"The authors have submitted an acceptable characterization of MSCs; however, the characterization data that include immunophenotyping and mesodermal differentiation would be complete when:

- chondrogenic differentiation data is included. A complete mesodermal differentiation based on acceptable standards comprises osteo, adipose and chondro differentiations;

"

Author response:

Analysis of the differentiation capability of MSCs into 3 mesodermal tissues according to the guidelines is actually needed. However, the completeness of the differentiation analysis is not mandatory. We refer to several articles related to the study of MSCs regarding the incompleteness of the differentiation analysis data:

3 1. Lotfinia M, Lak S, Mohammadi Ghahhari N, et al. Hypoxia Pre-Conditioned Embryonic Mesenchymal Stem Cell Secretome Reduces IL-10 Production by Peripheral Blood Mononuclear Cells. *Iran Biomed J*. 2017;21(1):24-31. doi:10.6091/21.1.24

9 2. Jeske R, Yuan X, Fu Q, Bunnell BA, Logan TM, Li Y. In Vitro Culture Expansion Shifts the

Immune Phenotype of Human Adipose-Derived Mesenchymal Stem Cells. *Front Immunol.*

2021;12:621744. Published 2021 Mar 10. doi:10.3389/fimmu.2021.621744

3. Qu X, Liu X, Cheng K, Yang R, Zhao RC. Mesenchymal stem cells inhibit Th17 cell differentiation by IL-10 secretion. *Exp Hematol.* 2012;40(9):761-770.

doi:10.1016/j.exphem.2012.05.006

However, the lack of MSCs differentiation data analysis remains a limitation of our study in which we will improve in further studies.

- "It has provided isotype or unstained control for lineage negative staining. As a practice, the negative population is usually located within the first quarter or first half of the antibody axis. In the current supplementary data, the negative population has been depicted in the center of the antibody axis."

Author response:

The isotype for lineage negative staining has been revised and included in the new extended data files. Reviewers could check our revised extended data on <https://doi.org/10.17605/OSF.IO/RJ8PC>

Competing Interests: The authors have no competing interests regarding this article.

Version 1

Reviewer Report 05 July 2021

<https://doi.org/10.5256/f1000research.54326.r87539>

© 2021 Moll G. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Guido Moll

¹ BIH Center for Regenerative Therapies (BCRT) and Berlin-Brandenburg School of Regenerative Therapies (BSRT), Berlin Institute of Health, Charité - University Medicine Berlin, Berlin, Germany

² BIH Center for Regenerative Therapies (BCRT) and Berlin-Brandenburg School of Regenerative Therapies (BSRT), Berlin Institute of Health, Charité - University Medicine Berlin, Berlin, Germany

³ BIH Center for Regenerative Therapies (BCRT) and Berlin-Brandenburg School of Regenerative Therapies (BSRT), Berlin Institute of Health, Charité - University Medicine Berlin, Berlin, Germany

In this Case Study, Putra *et al.* report the experimental treatment of three patients suffering from severe COVID-19 with hypoxia-conditioned mesenchymal stromal cell (MSC)-derived secretome (S-MSC) prepared by tangential flow filtration (TFF), to prepare the highly concentrated/enriched MSC secretome for clinical application. In their introduction and discussion, the authors identify among others the COVID-19 cytokine storm and concomitant severe lung and tissue damage, as targets of MSC therapy, anticipating that MSC therapy may exert pleiotropic beneficial effects.

1 The Case Study was conducted at Bethesda Hospital in Yogyakarta, Indonesia (With Local Health Research Committee approval number No.91/KEPK-RSB/V1/20), with written informed consent for the treatment with S-MSCs obtained from each patient prior to treatment. All patients received standard treatment for severe COVID-19, with on-top adjunct treatment with S-MSCs. This is a Case Report of three patients, who were all treated in exploratory fashion with different doses of S-MSCs, and there is no control group or control patients reported in the study.

The S-MSC was prepared by TFF to enrich bioactive factors secreted by the MSCs into 1ml therapeutic doses. The S-MSCs were applied in each of the three patients six times every 12 hours via deltoid intramuscular injection. Molecular weight cut-off combination of 10-50 kDa 50%, 50-100 kDa 25%, and 100-300 kDa 25% were used. The production method of S-MSC is outlined in reference number 12: Haraszti *et al.*¹ Exosomes Produced from 3D-cultures of MSCs by Tangential Flow Filtration Show higher Yield and Improved Activity published in Molecular Therapy in 2018.

The authors report:

- In Table 1) Clinical Characteristics of Patients Receiving S-MSC: Patient Sex, Age, Clinical Classification (Severe COVID-19), Principal Symptoms, Respective Comorbidities (e.g. Hypertension, T2D, Liver Failure, Stroke, Thalassemia);
- In Table 2) Standard Treatment Received by the Three Patients: Drugs Administered (Antiviral, Antibiotic, Antifungal, and Corticosteroid Treatment) and Oxygen Support (Before / After S-MSC Therapy);
- In Table 3) Comparison of Laboratory Parameters Before and After S-MSC Treatment (CRP, D-Dimer, Lymphocytes, SO2); and in Figure 1 and 2) Chest X-ray's of the Three Patients and a Schematic Study Overview, respectively.

Given the standard of care treatment, that there was no control group, and that this was an open label Case Study, the key findings of this report are as follows (although any of these findings could also be a result of the standard treatment):

1. CRP was strongly log-scale reduced after S-MSC treatment (from 62, 160, 118 to 0.4, 5.1, and 8.5),
2. D-dimer was reduced around 2-fold on average (1540, 880, 235 to 384, 660, 87),
3. Lymphocyte counts normalized/2-fold increased on average (15, 10, 12 to 20, 27, 26),
4. SO2 increased/improved (89.6, 90.6, 85 to 99.6, 95.9, 98) together with PO2, and 5) Chest x-rays mainly indicated that lung pathology and cardiomegaly improved (reduction of bilateral ground glass opacities and cardiomegaly).

In their introduction and discussion, the authors briefly outline the study hypothesis and why MSC / S-MSC treatment may be beneficial and relate this to the improved functional parameters in their Case Study, and these findings are in line with other clinical reports on MSC treatment of COVID-19. So it stands to reason that hypoxia conditioned S-MSC (concentrated soluble fraction of MSC media) is first of all safe, and second may be just as efficient as cellular MSCs in treating COVID-19. However, given the lack of a control group, and open-label treatment of these three patients the

findings should be interpreted with caution. Interestingly, not only inflammation and lung parameters improved, but also D-dimer levels, as typically observed in the follow-up of successfully treated patients, see "Viscoelastic Testing Reveals Normalization of the Coagulation Profile 12 Weeks After Severe COVID-19" ² published in *Scientific Reports* in 2021.

The authors recognized COVID-19 coagulopathy as a challenge for MSC therapy, as reported earlier in "MSC Therapies for COVID19: Importance of Patient Coagulopathy, Thromboprophylaxis, Cell Product Quality and Mode of Delivery for Treatment Safety and Efficacy" ³ published in *Frontiers in Immunology* 2020 and "Intravascular MSC Therapy Product Diversification: Time for New Clinical Guidelines" ⁴ published in *Trends in Molecular Medicine* 2019. Another crucial point is the appropriate timing and dosing and mode of application of MSC therapy in COVID-19, "Mesenchymal Stromal Cell Therapy for Coronavirus Disease 2019: Which? When? And How Much?" ⁵ published in *Cytotherapy* 2021.

The authors indicate their approach as a rescue treatment for patients progressed to severe COVID-19 stage (often presenting with pronounced coagulopathy). Here, the application of cellular secretome instead of the actual cellular MSC products may be somewhat superior/safer given the expression of varying levels of highly procoagulant Tissue Factor (TF/CD142) on MSCs (See PMID's 32574263 ³ and 30711482 ⁴). In this context, another key advantage of this study may be that the S-MSC was applied via deltoid intramuscular injection.

Future studies should also put more emphasis on well-controlled dose escalation, pharmacokinetics and pharmacodynamics, as well as biodistribution of the product with tight monitoring of coagulation/complement markers and other safety parameters, adjunct with deeper monitoring of a broader panel of biomarkers of response (e.g. systemic and potentially tissue cytokine profiling in blood and BAL as well as longitudinal monitoring of immune cell subsets in the periphery and local tissue BAL). Another important point is the timing of the treatment relative to the diseases stage (e.g. newly diagnosed vs. progressed patients, identify optimal treatment window). Presumably, S-MSC is an allogeneic off-the-shelf product, thus aspects of alloimmunization should be studied in more detailed (humoral and cellular alloimmunization). Another important issue is the GMP-compliant production of S-MSC for use in human patients.

References

1. Haraszti R, Miller R, Stoppato M, Sere Y, et al.: Exosomes Produced from 3D Cultures of MSCs by Tangential Flow Filtration Show Higher Yield and Improved Activity. *Molecular Therapy*. 2018; **26** (12): 2838-2847 [Publisher Full Text](#)
2. Magomedov A, Zickler D, Karaivanov S, Kurreck A, et al.: Viscoelastic testing reveals normalization of the coagulation profile 12 weeks after severe COVID-19. *Scientific Reports*. 2021; **11** (1). [Publisher Full Text](#)
3. Moll G, Drzeniek N, Kamhieh-Milz J, Geissler S, et al.: MSC Therapies for COVID-19: Importance of Patient Coagulopathy, Thromboprophylaxis, Cell Product Quality and Mode of Delivery for Treatment Safety and Efficacy. *Front Immunol*. 2020; **11**: 1091 [PubMed Abstract](#) | [Publisher Full Text](#)
4. Moll G, Ankrum JA, Kamhieh-Milz J, Bieback K, et al.: Intravascular Mesenchymal Stromal/Stem Cell Therapy Product Diversification: Time for New Clinical Guidelines. *Trends Mol Med*. **25** (2): 149-163 [PubMed Abstract](#) | [Publisher Full Text](#)
5. Shahani P, Datta I: Mesenchymal stromal cell therapy for coronavirus disease 2019: which? when? and how much?. *Cytotherapy*. 2021. [PubMed Abstract](#) | [Publisher Full Text](#)

Is the background of the cases' history and progression described in sufficient detail?

Yes

Are enough details provided of any physical examination and diagnostic tests, treatment given and outcomes?

Yes

Is sufficient discussion included of the importance of the findings and their relevance to future understanding of disease processes, diagnosis or treatment?

Yes

Is the conclusion balanced and justified on the basis of the findings?

Yes

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Immunology; Cardiovascular System; COVID-19; Stem Cell Therapy; Mesenchymal Stromal Cells (MSCs)

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard.

Author Response 17 Jul 2021

Agung Putra

Dear Reviewer

2 Thank you for giving us the opportunity to submit a revised of our manuscripts "Case series of the first three severe COVID-19 patients treated with the secretome of hypoxia-mesenchymal stem cells in Indonesia". We appreciate the time and effort that reviewers dedicated to providing feedback on our manuscript and are grateful for the insightful comments on and valuable improvements to our paper. We have incorporated most of the suggestions made by the reviewers. Those changes are highlighted within the manuscript and extended data. Please see below, in bold, for a point-by-point response to the reviewers' comments and concerns. All page numbers refer to the revised manuscript file with tracked changes (yellow highlight).

- This is a Case Report of three patients, who were all treated in exploratory fashion with different doses of S-MSCs, and there are no control group or control patients reported in the study.

Author response: Thank you for pointing this out. The reviewer is correct that we are reporting the findings of a case report study, and we do not utilize a control group to compare the relevance of the results.

- So, it stands to reason that hypoxia conditioned S-MSC (concentrated soluble fraction of MSC media) is first of all safe,

Author response: We confirm the reviewer's statement if certainly what was emphasized at the outset was the safety of the use of S-MSCs. We believe that S-MSC is safe because it has undergone a sterile ultrafiltration process using a TFF strategy of up to 10 kDa in size, so contamination is not possible. Future studies of S-MSCs will lead the safety parameters.

- Second maybe just as efficient as cellular MSCs in treating COVID-19.

Author Response: Regarding the efficacy of S-MSCs to treat the severe condition of COVID-19, we suggest that the S-MSCs is more efficient than cellular MSCs. This is due to the pulmonary intravascular coagulopathy (PIC) that present in severe COVID-19 patients could blockage the direction of cellular MSCs to homing to injured pulmonary. We use S-MSCs molecules under 300 kDa which makes it possible to get past the blockages due to PIC to modulate the inflammatory condition and enhance pulmonary regeneration.

- Interestingly, not only inflammation and lung parameters improved, but also D-dimer levels, as typically observed in the follow-up of successfully treated patients

Author response: We suggest that the possibility of S-MSCs to improve D-dimer levels, besides lung parameters is the controlled immune system's excessive inflammation and overactivation and the improvement of the lung caused by anti-inflammatory cytokines and growth factors contained in S-MSCs which possible to pass the PIC blockages.

- Future studies should also put more emphasis on well-controlled dose-escalation, pharmacokinetics, and pharmacodynamics, as well as biodistribution of the product with tight monitoring of coagulation/complement markers and other safety parameters, adjunct with deeper monitoring of a broader panel of biomarkers of response (e.g. systemic and potentially tissue cytokine profiling in blood and BAL as well as longitudinal monitoring of immune cell subsets in the periphery and local tissue BAL). Another important point is the timing of the treatment relative to the disease's stage (e.g. newly diagnosed vs. progressed patients, identify optimal treatment window).

Author response: We agree that this is a potential limitation of the study. In our future study, we will analyze the pharmacokinetics, pharmacodynamics, and biodistribution of the secretome. Furthermore, in the prospective study, we will also evaluate the parameter related to cytokine profiling in blood and BAL and monitoring of immune cell subsets in the periphery and local tissue BAL. We also agree that another potential limitation is no information about the disease stage of patients receiving secretome MSCs treatment. In the future study, it is planned to make several groups of patients who describe the different disease stages to determine the optimal treatment window.

- S-MSC is an allogeneic off-the-shelf product, thus aspects of alloimmunization should be studied in more detail (humoral and cellular alloimmunization). Another important issue is the GMP-compliant production of S-MSC for use in human patients

Author response: Thank you for pointing this out. We agree that this is an important

consideration, we have added the MSCs characterization in extended data, figure S1. Based on this data, the expression CD34, CD45, CD11b, CD19, and HLA-DR are represented as Lin <2%. It can be concluded that the possibility of an allergic reaction is quite limited. Although the potential of allergic reactions is limited, further studies need to be carried out.

Competing Interests: Authors declare that they have no conflict of interest.

Reviewer Report 05 July 2021

<https://doi.org/10.5256/f1000research.54326.r85411>

© 2021 Ramasamy R. This is an open access peer review report distributed under the terms of the [Creative Commons Attribution License](#), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

?

Rajesh Ramasamy 

¹ Department of Pathology, Universiti Putra Malaysia, Selangor, Malaysia

² Department of Pathology, Universiti Putra Malaysia, Selangor, Malaysia

³ Department of Pathology, Universiti Putra Malaysia, Selangor, Malaysia

The authors have tested the usefulness of mesenchymal stem/stromal cells-derived filtered supernatant to treat severe ARS due to COVID-19. The outcomes of the intervention were measured through the parameter of CRP, D-Dimer, and improvement of patients' clinical conditions. The tested treatment indeed showing a range of positive outcomes on the patients. The following technical queries require clarifications:

1. Provided the supplementary laboratory data on the characterisation of MSC.
2. Include the details of the hypoxic conditions that catered to the MSC.
3. What would be the rationale for selecting various percentages and molecular weight for filtering the supernatant?
4. Any profiling of cytokines or growth factors had been conducted prior to injecting the secretome into the patients?

The discussion should be made with achieved results rather than the anticipated immunosuppressive action of MSCs. There is no standardisation of MSC secretome. It could be possible that the exosome and microRNA in the secretome mediate the reparative function of MSCs through variable actions that include tissue repair.

Is the background of the cases' history and progression described in sufficient detail?

Yes

Are enough details provided of any physical examination and diagnostic tests, treatment

given and outcomes?

Partly

Is sufficient discussion included of the importance of the findings and their relevance to future understanding of disease processes, diagnosis or treatment?

Yes

Is the conclusion balanced and justified on the basis of the findings?

Partly

Competing Interests: No competing interests were disclosed.

Reviewer Expertise: Mesenchymal Stem Cell and Immunomodulation

I confirm that I have read this submission and believe that I have an appropriate level of expertise to confirm that it is of an acceptable scientific standard, however I have significant reservations, as outlined above.

Author Response 17 Jul 2021

Agung Putra

Dear Reviewer

2 Thank you for giving us the opportunity to submit a revised of our manuscripts "Case series of the first three severe COVID-19 patients treated with the secretome of hypoxia-mesenchymal stem cells in Indonesia". We appreciate the time and effort that reviewers dedicated to providing feedback on our manuscript and are grateful for the insightful comments on and valuable improvements to our paper. We have incorporated most of the suggestions made by the reviewers. Those changes are highlighted within the manuscript and extended data. Please see below, in bold, for a point-by-point response to the reviewers' comments and concerns. All page numbers refer to the revised manuscript file with tracked changes (yellow highlight).

- Provided supplementary laboratory data on the characterization of MSC.

Author response: Thank you for your suggestion. We have added the suggestion content on extended data, figure S1.

- Include the details of the hypoxic conditions that catered to the MSCs.

Author response: Thank you for this suggestion. We have added the details of hypoxic conditions on the discussion section that mark as the yellow highlight.

- What would be the rationale for selecting various percentages and molecular weight for filtering the supernatant?

Author response: We selected the 10-50 and 50-100 kDa molecular weight filtering to obtain the desired cytokines and growth factors, such as IL-10, TGF- β , VEGF, and PDGF. We also use 25% S-MSCs with 100-300 kDa categories to obtain the miRNA and

exosome. However, we do not analyze the exosome and miRNA contained in S-MSCs. Further studies need to be carried out.

- Any profiling of cytokines or growth factors had been conducted prior to injecting the secretome into the patients?

Author response: We have evaluated the cytokine and growth factor profiles before injecting secretome MSCs into patients. We are confirming that the quality of the secretome MSCs administered is consistent on all three patients (See extended data, table S1 for more details).

- The discussion should be made with achieved results rather than the anticipated immunosuppressive action of MSCs. There is no standardisation of MSC secretome. It could be possible that the exosome and microRNA in the secretome mediate the reparative function of MSCs through variable actions that include tissue repair.

Author response: Thank you for your suggestion. Accordingly, throughout the manuscript, we have revised and discussed the results in the discussion section such as:

“The clinical symptoms were significantly improved with patchy GGO improvement, associated with the decrease of D-dimer and CRP level and increase of SO 2 and PO 2. From our observation, we would suggest that six doses of S-MSCs performed the most optimal treatment in the COVID-19 patients. These results suggest that the immune system's excessive inflammation and overactivation were alleviated by anti-inflammatory cytokines contained in S-MSCs, while the high level of growth factors in S-MSCs could also accelerate the improvement of GGOs.”

“In our observations, the level of CRP and D-dimer was decreased in all patients treated with S-MSCs. This data suggest that S-MSCs could effectively control the overactivated immune cells. A previous study reported that S-MSCs could control proinflammatory immune cells due to their anti-inflammatory cytokines, such as IL-10 and TGF- β ^{11, 17}.”

We also agree with the reviewer's assessment that there is no standardization of secretome MSCs. Secretome MSCs contain exosome and soluble molecule active. This is a potential limitation of the study because we have not standardized the secretome MSCs. Therefore, further studies of secretome MSCs standardization need to be carried out.

Competing Interests: Authors declare that they have no conflict of interest.

The benefits of publishing with F1000Research:

- Your article is published within days, with no editorial bias
- You can publish traditional articles, null/negative results, case reports, data notes and more
- The peer review process is transparent and collaborative
- Your article is indexed in PubMed after passing peer review
- Dedicated customer support at every stage

For pre-submission enquiries, contact research@f1000.com

 F1000Research